

# PEGylated Molecule Assay Kit Technical Manual No. TM0644

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| I.    | Description                                    | 1  |
|-------|--|----|
| II.   | Kit Content                                    | 2  |
| III.  | Storage  | .2 |
| IV.   | Reagents/Equipments Required But Not Supplied  | 3  |
| ۷.    | Protocol                                       | 3  |
|       | 1. Sample Preparation                          | 3  |
|       | 2. Reagent Preparation                         | 3  |
|       | 3.Test Procedure                               | 4  |
|       | PEGylated Molecule / Biotin-PEG mAb Incubation | 4  |
|       | Streptavidin-HRP Incubation                    | 5  |
|       | Substrate Reaction and Absorbance Measurement  | 5  |
|       | 4. Calculation of Data                         | 5  |
| VI.   | Assay Procedure Summary                        | 6  |
| VII.  | Typical Assy Data                              | 7  |
| VIII. | Precision                                      | 8  |
| IX.   | Sensitivity                                    | 8  |
| Х.    | Recovery                                       | 8  |
| XI.   | Linearity                                      | 10 |
| XII.  | Interference                                   | 12 |
| XIII. | Pharmacokinetic Study                          | 13 |
| XIV.  | Troubleshooting                                | 14 |
| XV.   | Related Products1                              | 15 |
| XVI.  | Plate Layout                                   | 15 |

# **I. DESCRIPTION**

PEG (Polyethylene glycol) is a polyether compound with many applications from industrial manufacturing to medicine. PEGylation is a technology that covalently couples non-toxic, hydrophilic PEG to a drug. It is an FDA-approved method for the delivery of protein drugs. PEG modification can reduce the drug immunogenicity and antigenicity. The PEGylated drug decelerates renal excretion, improves stability towards proteolysis and increases its half life in blood. Accurate and sensitive quantification of PEGylated molecules is important for PEG conjugated product development and pharmaceutical study. PEG Antibody is a useful tool for the detection of PEGylated molecules.

**GenScript PEGylated Molecule Assay Kit,** a 1.5 hour competitive ELISA Kit, is developed for rapid detection of PEGylated molecules. This kit is based on competitive ELISA method. PEG Molecule Plate, which comprises of 8 wells x 12 strips, is demountable. It is a 96-well microtiter plate coated with PEGylated molecule. When Biotin-PEG mAb and PEGylated molecule are added to the well, the PEGylated molecule coated on the plate compete with the PEGylated molecule in the solution to interact with Biotin-PEG mAb. PEG standard control provided in the kit can be used for semiquantitative measurement of PEGylated molecules. To quantitate a PEGylated molecule of interest, operator should use individual PEGylated



molecule of interest to establish standard curve. (Principle is showed as below.)



Streptavidin-HRP is used for enzyme reaction to develop signal. In the test, PEGylated molecule, Biotin-PEG mAb and Streptavidin-HRP form a complex in the wells on the plate. Other unbound molecules can be removed by wash solution. The Streptavidin-HRP reacts with TMB substrate to develop blue product in the solution. The reaction is stopped by adding stop solution and the color turns yellow which can be read at 450 nm by Microtiter plate reader. Each absorbance value is correlated to each PEGylated molecule concentration in solution. The signal is inversely proportional to the PEGylated molecule concentration in the sample. PEGylated molecule standards of known concentration and the corresponding absorbance values are used to construct a standard curve. With the standard curve, PEGylated molecule amount present in the unknown sample can be calculated by transforming its absorbance value.

## **II. KIT CONTENT**

| Component Quantity              |                               | Part. No |
|---------------------------------|-------------------------------|----------|
| PEG Molecule Plate              | 1 plate (8 wells x 12 strips) | 458-80   |
| Assay Diluent                   | 60 ml                         | 458-60   |
| Biotin-PEG mAb Stock            | 100 µl                        | 458-20   |
| Antibody Dilution Buffer        | 10 ml                         | 458-90   |
| Streptavidin-HRP                | 12 ml                         | 458-30   |
| PEG Standard Control (10 µg/ml) | 50 µl                         | 458-10   |
| 20 × Wash Solution              | 40 ml                         | 458-70   |
| TMB Substrate                   | 12 ml                         | 458-40   |
| Stop Solution                   | 6 ml                          | 458-50   |
| Plate Sealer                    | 2                             | N/A      |
| User Manual                     | 1                             | N/A      |

The kit provides all reagents and solutions for PEGylated molecule detection.

# III. STORAGE

The unopened kit is stable for at least 12 months if stored at 2-8 °C and the opened kit is stable for up to 2 weeks at 2-8 °C. Do not freeze the kit.





# IV. REAGENTS/EQUIPMENTS REQUIRED BUT NOT SUPPLIED

Well characterized PEGylated molecule of interest of known concentration to prepare the standard Microtiter plate reader capable of measuring absorbance at 450 nm Automated microplate washer to wash the plate Deionized or distilled water to dilute 20 x Wash Solution Graduated cylinder to prepare Wash Solution Plastic container to store Wash Solution Tubes to aliquot and dilute samples Precision pipettes to deliver 10 µl, 100 µl, 200 µl and 1000 µl content 10 µl, 100 µl, 200 µl and 1000 µl pipette tips Multichannel pipettor Disposable reagent reservoir Paper towel Laboratory timer Refrigerator to store samples and kit components

## **V.PROTOCOL**

- All reagents in the kit and test samples should be equilibrated to room temperature before test.
- Preliminary experiment should be performed to optimize the sample dilution.

#### 1. Sample Preparation

- Handle serum and plasma samples in accordance with NCCLS (National Committee for Clinical Laboratory Standards) guidelines for preventing transmission of blood-borne infection.
- Assay Diluent is used for sample dilution.
- For serum and plasma sample containing high level of PEGylated molecule, a minimum of 1:100 dilution without heat treatment is recommended to remove matrix interference in the assay.
- For serum and plasma sample containing low level of PEGylated molecule, heat treatment is recommened. A minimum of 1:20 dilution is needed and the diluted samples should be boiled for 5 minutes to remove matrix interference in the assay. Don't boil the sample for a long time to avoid sample getting destroyed.
- The results showed samples with heat treatment have better recovery than those with 1:100 dilution without heat treatment.

**Serum:** Use a blood separator tube and allow the sample to clot for 30 minutes. Centrifuge for 10 minutes at 1000 x g. Run the assay immediately, otherwise aliquot and store the sample below -20 °C. Avoid repeated thaw-freeze cycle.

**Plasma:** Treat the blood with citrate, EDTA or heparin as an anticoagulant. Centrifuge for 10 minutes at 1000 x g within 30 minutes for plasma collection. Run the assay immediately. Otherwise aliquot and store the sample below -20 °C. Avoid repeated thaw-freeze cycle.

**Cell culture supernatant:** Centrifuge the sample to remove the particulate materials. Run the assay immediately, otherwise aliquot and store sample below -20 °C. Avoid repeat thaw-freeze cycle.

#### 2. Reagent Preparation

• If any precipitate is found in the 20 × Wash Solution, incubate the bottle in water bath (up to 50 °C) with occasional mixing until all the precipitate is dissolved.



**1 x Wash Solution:** Dilute  $20 \times Wash$  Solution by 1:19 v/v with deionized or distilled water. For example, dilute 40 ml of  $20 \times Wash$  Solution with 760 ml of deionized or distilled water to make 800 ml of 1 × Wash Solution. Store at 2-8 °C.

**Biotin-PEG mAb Solution:** Dilute Biotin-PEG mAb Stock by 1:100 (v/v) with Antibody Dilution Buffer.

#### **PEG Molecule Standards Preparation**

To detect PEGylated molecule amount precisely, operator should establish a standard curve with the PEGylated molecule of interest of known concentration. The standard curve established by PEGylated molecule can be used to quantitate the same PEG molecule and is semiquantitative for other PEGylated molecules. Peginterferon Alfa 2A standard curve was established with Peginterferon Alfa 2A as standards in <u>pharmacokinetic study</u> section. It can serve as a typical example for individual PEGylated protein quantitation. Peginterferon Alfa 2A was the protein drug pegylated with a branched 40 kDa PEG chain, which is produced by Roche pharmaceutical company.

• Allow PEG Standard Control to sit for a minimum of 15 minutes at room temparature with gentle agitation prior to making dilution.

- 2.1 Label seven 1.5 ml Eppendorf tubes with '81 ng/ml', '27 ng/ml', '9 ng/ml', '3 ng/ml', '1 ng/ml', '0.333 ng/ml' and '0 ng/ml'.
- 2.2 Pipette 8.1 µl of *PEG Standard Control* and 991.9 µl of *Sample Diluent* into the tube labeled with '81 ng/ml' and vortex it.
- 2.3 Pipette 400 µl of Sample Diluent into the rest empty tubes.
- 2.4 Pipette 200 μl of 81 ng/ml of PEG molecule solution to the tube labeled with '27 ng/ml' and vortex it to make the standard point of 27 ng/ml.
- 2.5 Similarly, prepare the rest standard series (9, 3, 1, 0.333 ng/ml).

#### **PEG Molecule Plate Preparation**

- It is recommended that all PEG standards and samples should be prepared in duplicate.
- Count the strips for the assay and make sure the strips are tightly snapped in the plate frame.
- Leave the unused strips in the foil pouch and store at 2-8 °C. The strips must be stored in the closed foil pouch to prevent moisture because the moisture can damage the PEGylated Molecule Plate.

## 3. Test Procedure

• Pat the plate on paper towel to remove residual liquid in the wells after wash step.

#### PEGylated Molecule / Biotin-PEG mAb Incubation

- 3.1 Add 100 µl of *Assay Diluent* to the NSB (Non-specific binding) wells and 50 µl of a set of prepared PEG standards or samples to the remaining wells separately.
- 3.2 Add 50 µl of prepared *Biotin-PEG mAb* to each well except the NSB wells.
- 3.3 Cover the plate with *Plate Sealer* and incubate at 25 °C for 60 minutes.
- 3.4 Wash the plate with 260  $\mu$ l of 1 x Wash Solution for four times.



#### **Streptavidin-HRP Incubation**

- 3.5 Add 100 µl of Streptavidin-HRP to all the wells.
- 3.6 Cover the plate with *Plate Sealer* and incubate at 37 °C for 10 minutes.
- 3.7 Wash the plate with 260  $\mu$ l of 1 x Wash Solution for four times.

### **Substrate Reaction and Absorbance Measurement**

- 3.8 Add 100 µl of *TMB Substrate* to all the wells and incubate at 25 °C for 15-20 minutes and protect it from the light.
- 3.9 Add 50 µl of Stop Solution to all the wells to stop the enzyme reaction.
- 3.10 Read the plate on Microtiter plate reader at 450 nm.

## 4. Calculation of Data

- To ensure test stability, read the plate at 450 nm immediately after Stop Solution addition.
- If the sample is diluted, multiply the interpolated value by the dilution factor to calculate the amount of PEGylated molecule in sample.
- 4.1 Calculate the average optical density (OD) for each set of replicate wells.
- 4.2 Calculate the adjusted average OD by subtracting the average NSB OD from the average OD for each standard and sample.

Adjusted average OD = Average OD – Average NSB OD

4.3 Calculate the percent bound for each standard and sample using the following relationship

$$B/B_0 (\%) = (Standard or sample OD - NSB OD) X 100$$
(Zero standard OD - NSB OD)

- 4.4 Generate a standard curve by plotting the B/B<sub>0</sub> on the vertical (Y) axis versus the PEGylated molecule standard concentration on the horizontal (X) axis.
- 4.5 The amount of PEGylated molecule in sample is determined by extrapolating its OD value to the standard curve.

5



# VI. ASSAY PROCEDURE SUMMARY



6



# **VII. TYPICAL ASSAY DATA**

The standard curve below was provided for demonstration only. Operator should use PEGylated molecule of interest to set up standard curve to precisely determine individual PEGylated molecule each time.

| DEO Otan dand           |             | OD          | 450     |                     |                   |
|-------------------------|-------------|-------------|---------|---------------------|-------------------|
| PEG Standard<br>(ng/ml) | Duplicate 1 | Duplicate 2 | Average | Adjusted<br>Average | %В/В <sub>о</sub> |
| NSB                     | 0.059       | 0.058       | 0.059   | -                   | -                 |
| 0                       | 1.995       | 1.988       | 1.992   | 1.933               | -                 |
| 0.333                   | 1.665       | 1.679       | 1.672   | 1.613               | 84                |
| 1                       | 1.030       | 0.992       | 1.011   | 0.952               | 49                |
| 3                       | 0.445       | 0.437       | 0.441   | 0.382               | 20                |
| 9                       | 0.274       | 0.252       | 0.263   | 0.204               | 11                |
| 27                      | 0.197       | 0.181       | 0.189   | 0.130               | 7                 |
| 81                      | 0.139       | 0.130       | 0.135   | 0.076               | 4                 |





## VIII. PRECISION

**Intra-assay:** Three different levels of PEG40 kDa were tested 20 times on one plate to assess intra-assay precision.

**Inter-assay:** Three different levels of PEG40 kDa were tested in 20 separate assays to assess inter-assay precision.

| Intra-assay            |                 |       |       |                        | Inter-          | assay |       |
|------------------------|-----------------|-------|-------|------------------------|-----------------|-------|-------|
| # of<br>replicate<br>s | Mean<br>(ng/ml) | SD    | CV%   | # of<br>replicate<br>s | Mean<br>(ng/ml) | SD    | CV%   |
| 20                     | 0.556           | 0.056 | 10.1% | 20                     | 0.480           | 0.034 | 7.0%  |
| 20                     | 0.903           | 0.101 | 11.2% | 20                     | 0.905           | 0.053 | 5.8%  |
| 20                     | 2.922           | 0.337 | 11.5% | 20                     | 2.815           | 0.331 | 11.8% |

## IX. SENSITIVITY

The Biotin-PEG mAb in the kit is specific to the backbone of PEG and it is able to bind a broad variety of PEG molecules. Several PEG molecules with different molecular weight have been tested in the assay. The IC50 of each PEG molecule was presented as below.

| PEG       | IC50 (ng/ml) | PEG             | IC50 (ng/ml) |
|-----------|--------------|-----------------|--------------|
| PEG40 kDa | 0.947        | PEG5 kDa        | 568.6        |
| PEG30 kDa | 0.844        | PEG1 kDa        | 469500       |
| PEG20 kDa | 0.861        | 8arm-PEG-40 kDa | 1.3          |
| PEG10 kDa | 37.5         | 4arm-PEG-10 kDa | 1150.3       |

## X. RECOVERY

#### Samples with heat treatment

PEG40 kDa was spiked separately into 1:20 diluted matrices throughout the range of the assay and then were boiled for 5 minutes. The percentage was determined by dividing the calculated value of each spiked sample by the corresponding spiked concentration.

| Sample             | Spike (ng/ml) | Mean Recovery (%) | Range (%) |
|--------------------|---------------|-------------------|-----------|
|                    | 1             | 95                | 91-98     |
| Mouse Serum (n=5)  | 3             | 106               | 95-121    |
|                    | 6             | 95                | 83-104    |
|                    | 1             | 96                | 90-106    |
| Mouse Plasma (n=5) | 3             | 100               | 94-108    |
|                    | 6             | 102               | 89-109    |
|                    | 1             | 101               | 96-106    |
| Rat Serum (n=5)    | 3             | 93                | 90-103    |
|                    | 6             | 86                | 67-108    |

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|                    | 1 | 98  | 95-102  |
|--------------------|---|-----|---------|
| Rat Plasma (n=5)   | 3 | 104 | 103-105 |
|                    | 6 | 108 | 94-117  |
|                    | 1 | 102 | 96-111  |
| Human Serum (n=5)  | 3 | 102 | 94-109  |
|                    | 6 | 98  | 87-106  |
|                    | 1 | 100 | 94-105  |
| Human Plasma (n=5) | 3 | 98  | 96-102  |
|                    | 6 | 99  | 86-108  |

## Samples with high dilution

PEG40 kDa was spiked separately into 1:100 diluted matrices throughout the range of the assay without heat treatment. The percentage was determined by dividing the calculated value of each spiked sample by the corresponding spiked concentration.

| Sample             | Spike (ng/ml) | Mean Recover (%) | Range (%) |
|--------------------|---------------|------------------|-----------|
|                    | 1             | 98               | 91-100    |
| Mouse Serum (n=5)  | 3             | 96               | 85-110    |
|                    | 6             | 91               | 82-103    |
|                    | 1             | 103              | 97-108    |
| Mouse Plasma (n=5) | 3             | 103              | 95-110    |
|                    | 6             | 99               | 89-108    |
|                    | 1             | 101              | 90-108    |
| Rat Serum (n=5)    | 3             | 99               | 96-105    |
|                    | 6             | 98               | 88-107    |
|                    | 1             | 99               | 83-107    |
| Rat Plasma (n=5)   | 3             | 97               | 89-100    |
|                    | 6             | 98               | 91-105    |
|                    | 1             | 102              | 98-102    |
| Human Serum (n=5)  | 3             | 102              | 92-108    |
|                    | 6             | 101              | 88-115    |
|                    | 1             | 103              | 94-109    |
| Human Plasma (n=5) | 3             | 107              | 101-113   |
|                    | 6             | 96               | 82-105    |



# XI. LINEARITY

## Samples with heat treatment

PEG40 kDa was spiked into 1:20 diluted matrices at the concentration of 4 ng/ml and then were boiled for 5 minutes. The prepared samples were serially diluted in the Assay Diluent. The percentage was determined by dividing the calculated concentration of each diluted sample by the corresponding spiked concentration.

| Dilution |                       | Mouse Serum (n=5) | Mouse Plasma (n=5) |
|----------|-----------------------|-------------------|--------------------|
| 1:2      | Average % of Expected | 103               | 107                |
|          | Range (%)             | 99-106            | 100-115            |
| 1.1      | Average % of Expected | 102               | 98                 |
| 1.4      | Range (%)             | 94-108            | 95-103             |
| 1.0      | Average % of Expected | 96                | 86                 |
| 1:8      | Range (%)             | 91-101            | 83-90              |

| Dilution |                       | Rat Serum (n=5) | Rat Plasma (n=5) |
|----------|-----------------------|-----------------|------------------|
| 1:2      | Average % of Expected | 104             | 102              |
|          | Range (%)             | 96-110          | 98-109           |
| 1:4      | Average % of Expected | 103             | 93               |
|          | Range (%)             | 99-109          | 88-97            |
| 1.0      | Average % of Expected | 96              | 95               |
| 1:8      | Range (%)             | 85-105          | 90-101           |

| Dilution |                       | Human Serum (n=5) | Human Plasma (n=5) |
|----------|-----------------------|-------------------|--------------------|
| 1.0      | Average % of Expected | 105               | 95                 |
| 1:2      | Range (%)             | 93-114            | 91-103             |
| 1.1      | Average % of Expected | 103               | 96                 |
| 1.4      | Range (%)             | 100-106           | 89-101             |
| 1.0      | Average % of Expected | 101               | 87                 |
| 1.8      | Range (%)             | 91-107            | 81-93              |

 10

 860 Centennial Ave., Piscataway, NJ 08854, USA

 Toll-Free: 1-877-436-7274

 Tel: 1-732-885-9188

 Fax: 1-732-210-0262

 Email: product@genscript.com

 Web: www.genscript.com



## Samples with high dilution

PEG40 kDa was spiked into 1:50 diluted matrices at the concentration of 8 ng/ml without heat treatment. The prepared samples were serially diluted in the Assay Diluent. The percentage was determined by dividing the calculated concentration of each diluted sample by the corresponding spiked concentration.

| Dilution |                       | Mouse Serum (n=5) | Mouse Plasma (n=5) |
|----------|-----------------------|-------------------|--------------------|
| 1.0      | Average % of Expected | 89                | 101                |
| 1.2      | Range (%)             | 84-99             | 95-106             |
| 1.1      | Average % of Expected | 101               | 108                |
| 1.4      | Range (%)             | 92-107            | 103-118            |
| 1.0      | Average % of Expected | 106               | 101                |
| 1.0      | Range (%)             | 97-112            | 96-106             |

| Dilution |                       | Rat Serum (n=5) | Rat Plasma (n=5) |  |  |
|----------|-----------------------|-----------------|------------------|--|--|
| 1:2      | Average % of Expected | 100             | 96               |  |  |
|          | Range (%)             | 84-111          | 90-99            |  |  |
| 1:4      | Average % of Expected | 105             | 104              |  |  |
|          | Range (%)             | 89-115          | 94-112           |  |  |
| 1:8      | Average % of Expected | 99              | 98               |  |  |
|          | Range (%)             | 93-106          | 91-107           |  |  |

| Dilution |                       | Human Serum (n=5) | Human Plasma (n=5) |  |
|----------|-----------------------|-------------------|--------------------|--|
| 1:2      | Average % of Expected | 95                | 99                 |  |
|          | Range (%)             | 85-103            | 94-102             |  |
| 1:4      | Average % of Expected | 108               | 104                |  |
|          | Range (%)             | 106-110           | 95-112             |  |
| 1:8      | Average % of Expected | 109               | 98                 |  |
|          | Range (%)             | 104-112           | 96-102             |  |



## XII. INTERFERENCE

Some reagents such as Tween, Triton and NP-40 interfere with test results due to structural similarity to PEG. Compatibility of tissue/cell homogenization buffers should be tested before the assay. SDS and RIPA buffer was added into *Assay Diluent* with serial percentage. Same amount of PEG molecules were spiked into the buffers to evaluate reagent interference. The test result demonstrated that RIPA cell lysis buffer should be used below 0.025% (v/v) and SDS is compatible at or below 0.0125% (m/v) in the assay.

| <br>Percent (%) | SDS     | RIPA    |  |  |
|-----------------|---------|---------|--|--|
| <br>0.1         | 69.10%  | 21.90%  |  |  |
| 0.05            | 78.20%  | 28.10%  |  |  |
| 0.025           | 90.80%  | 58.70%  |  |  |
| 0.0125          | 97.90%  | 92.30%  |  |  |
| 0.00625         | 99.10%  | 98.30%  |  |  |
| 0.003125        | 101.80% | 102.90% |  |  |
| 0.001563        | 105.20% | 105.20% |  |  |
|                 |         |         |  |  |

The variety of PEGylated molecules were spiked in Assay Diluent at different concentrations. One set of the samples were treated with heat treatment which was boiled at 100 °C for 5 minutes. The samples with heat treatment and normal samples were assayed in one plate at the same time. The comparison of test results is listed as below which indicates that the heat treatment for the test samples is acceptable in the assay.

| PEGylated molecules   | ng/ml | No Treatment | Heat Treatment |  |
|-----------------------|-------|--------------|----------------|--|
|                       | 0     | 2.260        | 2.115          |  |
|                       | 0.333 | 1.901        | 1.946          |  |
| PEG40 KDa             | 3     | 0.529        | 0.584          |  |
|                       | 27    | 0.203        | 0.216          |  |
|                       | 0     | 2.251        | 2.131          |  |
|                       | 0.333 | 1.985        | 1.942          |  |
| PEG40 KDa-BSA         | 3     | 0.912        | 0.894          |  |
|                       | 27    | 0.283        | 0.285          |  |
|                       | 0     | 2.171        | 2.115          |  |
|                       | 0.333 | 1.927        | 1.960          |  |
| PEG40 KDa-1gG         | 3     | 0.774        | 0.761          |  |
|                       | 27    | 0.259        | 0.251          |  |
|                       | 0     | 2.280        | 2.192          |  |
| Peginterferon Alfa 2A | 0.037 | 1.998        | 1.973          |  |
|                       | 0.333 | 1.263        | 1.282          |  |
|                       | 3     | 0.319        | 0.317          |  |



# XIII. PHARMACOKINETIC STUDY

Peginterferon Alfa 2A was used as standard PEGylated molecule to set up standard curve. Peginterferon Alfa 2A is a PEGylated drug. The rabbit was injected with Peginterferon Alfa 2A at the dose of 8.40 µg/kg. The animal blood was collected at different time points. With the standard curve, pharmacokinetic study was performed to measure the PEGylated drug concentration in blood. The standard curve and drug concentration was listed as below.

#### Peginterferon Alfa 2A standard curve



| Peginterferon Alfa | Dunlicate 1 | Duplicate | Adjusted | B/B <sub>0</sub> (%) |  |
|--------------------|-------------|-----------|----------|----------------------|--|
| 2/ ( (iig/iiii)    | Duplicate 1 | 2         | Average  |                      |  |
| NSB                | 0.056       | 0.059     | 0.058    | -                    |  |
| 0                  | 1.959       | 1.870     | 1.857    | -                    |  |
| 0.037              | 1.820       | 1.755     | 1.730    | 93                   |  |
| 0.111              | 1.613       | 1.551     | 1.524    | 82                   |  |
| 0.333              | 1.248       | 1.186     | 1.159    | 62                   |  |
| 1                  | 0.517       | 0.558     | 0.48     | 26                   |  |
| 3                  | 0.176       | 0.196     | 0.128    | 7                    |  |
| » 9                | 0.136       | 0.127     | 0.074    | 4                    |  |

# Test Result of Peginterferon Alfa 2A concentration in rabbit blood



| Time (h) | Peginterferon Alfa |  |  |  |  |
|----------|--------------------|--|--|--|--|
|          | ZA (ng/mi)         |  |  |  |  |
| 0        | 0                  |  |  |  |  |
| 0.5      | 1.46               |  |  |  |  |
| 1.5      | 5.18               |  |  |  |  |
| 3        | 14.33              |  |  |  |  |
| 6        | 25.20              |  |  |  |  |
| 24       | 49.14              |  |  |  |  |
| 48       | 57.05              |  |  |  |  |
| 72       | 71.58              |  |  |  |  |
| 96       | 55.10              |  |  |  |  |
| 120      | 33.13              |  |  |  |  |
| 144      | 22.35              |  |  |  |  |
| 168      | 15.79              |  |  |  |  |

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# XIV.TROUBLESHOOTING

| Problem         | Probable Cause   | Solution  |  |  |  |
|-----------------|--|---|--|--|--|
|                 | Wells are not washed or aspirated properly                     | Make sure the wash apparatus work properly and wells are dry after aspiration               |  |  |  |
|                 | Wells are scratched with pipette tip or washing needles.       | Dispense and aspirate solution into and out of wells with caution                           |  |  |  |
| Poor Precision  | Particulates are found in the samples                          | Remove any particulates by centrifugation prior to the assay                                |  |  |  |
|                 | Serum is turbid after heat treatment                           | Use 30-fold dilution with sample diluent and/or centrifuge the treated sample at high speed |  |  |  |
|                 | Hemolysis in blood sample                                      | Prepare new samples   |  |  |  |
|                 | Improper preparation of standards                              | Prepare new standards as the manual described   |  |  |  |
|                 | Wells are not washed or aspirated properly                     | Make sure the wash apparatus work properly and wells are dry after aspiration               |  |  |  |
| Door Standard   | Pipetting error  | Check pipette calibration and repeat assay  |  |  |  |
| Curve           | Components are used from other lots or sources                 | Never substitute any components from another kit  |  |  |  |
|                 | Components are not brought to room                             | Repeat assay with components that have been   |  |  |  |
|                 | temperature prior to assay                                     | equilibrated to room temperature  |  |  |  |
|                 | Incubation steps are performed at wrong temperatures           | Perform incubation step as the manual describes   |  |  |  |
|                 | TMB substrate are not added or added at the wrong time         | Follow the manual to add the substrate  |  |  |  |
|                 | Streptavidin-HRP are not added, or added at the wrong time     | Follow the manual to repeat the assay   |  |  |  |
|                 | Components are used from other lots or sources                 | Use only lot-specific components  |  |  |  |
| Weak/No Signal  | TMB substrate is contaminated                                  | Use new TMB substrate   |  |  |  |
|                 | Do not add the proper volumes of reagents                      | Repeat assay with the required volumes in manual  |  |  |  |
|                 | Do not incubate the plate for proper time or temperature       | Follow the manual to repeat assay   |  |  |  |
|                 | Do not read the plate immediately after stop solution is added | Read the plate within 30 minutes  |  |  |  |
|                 | Plate is not washed properly                                   | Make sure the wash apparatus can work properly  |  |  |  |
|                 | TMB substrate is contaminated                                  | Use new TMB substrate with same Lot   |  |  |  |
| High Background | Evaporation of wells during incubations                        | Perform incubation steps with plate sealer in repeat assay                                  |  |  |  |
|                 | Incorrect incubation times and/or temperatures                 | Follow the manual to repeat the assay   |  |  |  |
|                 | TMB substrate is exposed to light                              | Use new TMB substrate   |  |  |  |



# **XV. RELATED PRODUCTS**

| <ul> <li>THE<sup>™</sup> PEG Antibody, mAb, Mouse</li> </ul> | A01795-100 |
|--|------------|
| ● THE <sup>™</sup> PEG Antibody [Biotin], mAb, Mouse         | A01796-100 |
| Protein A CIP Resin  | L00433     |
| Streptavidin-HRP   | M00091     |
| <ul> <li>Protein A Antibody, mAb, Mouse</li> </ul>           | A01778     |
| <ul> <li>Protein A Antibody [Biotin], mAb, Mouse</li> </ul>  | A01779     |
| <ul> <li>Protein A Antibody, pAb, Chicken</li> </ul>         | A00728     |
| <ul> <li>Protein A Antibody [HRP], pAb, Chicken</li> </ul>   | A00729     |
| Protein A ELISA Kit  | L00430     |

# **XVI. PLATE LAYOUT**

Use this plate layout to record standards and samples assayed.

|   | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|---|---|---|---|---|---|---|---|---|----|----|----|
| Α |   |   |   |   |   |   |   |   |   |    |    |    |
| в |   |   |   |   |   |   |   |   |   |    |    |    |
| С |   |   |   |   |   |   |   |   |   |    |    |    |
| D |   |   |   |   |   |   |   |   |   |    |    |    |
| Е |   |   |   |   |   |   |   |   |   |    |    |    |
| F |   |   |   |   |   |   |   |   |   |    |    |    |
| G |   |   |   |   |   |   |   |   |   |    |    |    |
| н |   |   |   |   |   |   |   |   |   |    |    |    |

#### Notes:

- Use this plate layout to record standards and samples assayed.
- The product is used for detection of PEGylated molecules in serum, plasma and other biological samples. The operator should read technical manual carefully before using this product.
- For research use only. Not for diagnostic use.

#### GenScript USA Inc.

860 Centennial Ave., Piscataway, NJ 08854 Tel: 732-885-9188, 732-885-9688 Fax: 732-210-0262, 732-885-5878 Email: product@genscript.com Web: www.genscript.com **For Research Use Only**